**International Journal of Pharmaceutics and Drug Research** 

 **ISSN: 2347-6346 Available online at http://ijpdr.com**

**Original Research Article**

**STUDY OF HISTAMINERGIC TRANSMISSION DURING SOCIAL INTERACTION IN** 

#### **MICE**

# **Ashish Pal\*, Lokesh Verma, Jitendra Banweer SOCIAL INTERAC<br>
MICE**<br> **Ashish Pal\*, Lokesh Verma, Jitendra Banweer<br>
Sagar Institute of Research & Technology-Pharmacy, Bhopal (M.P.)**

**ABSTRACT**



Social interaction is a fundamental aspect of behavior in many species, including mice, where it plays a crucial role in various social behaviors such as mating, territoriality, and social bonding (Yoshikawa *et al.,*  2006). Understanding the neural underlying social interaction is of great interest, not only for unraveling the complexities of social behavior but also for shedding light on potential therapeutic interventions for social deficits observed in neuropsychiatric disorders such as autism spectrum disorders (ASD) and schizophrenia. mechanisms

Histamine, a neurotransmitter primarily known for its role in regulating allergic implicated in modulating social behaviors (Passani & Blandina, 2011). The histaminergic system in the brain originates from neurons located in the tuberomammillary nucleus of the hypothalamus and projects widely throughout the central nervous system (Lee *et al.,* 2005). Mounting evidence suggests that histamine Mounting evidence suggests that histamine<br>may play a pivotal role in regulating social behavior, yet the precise mechanisms by which histaminergic transmission influences social interaction remain to be fully elucidated. Several studies have highlighted the involvement of histamine in social behaviors. the precise mechanisms<br>istaminergic transmission influer<br>interaction remain to be f<br>ed. Several studies have highligh<br>olvement of histamine in so

For instance, pharmacological studies utilizing histamine receptor agonists and antagonists have demonstrated alterations in social behavior in rodents (Takahashi *et al.,*  2006). Additionally, genetic studies in mice with manipulations of histamine receptors or histamine synthesis enzymes have provided further evidence of the importance of histamine signaling in social interaction (Yoshikawa *et al.,* 2006).

Despite these advances, there is still a need for comprehensive investigations into the specific neural circuits and molecular mechanisms through which histaminergic transmission modulates social behavior (Passani & Blandina, 2011). Moreover, the majority of existing studies have focused on the role of histamine in either aggressive or affiliative aspects of social behavior, with less attention given to the dynamic changes in histaminergic activity during social interaction.

In this study, aim to investigate the role of histaminergic transmission during social interaction in mice. Firstly, we will assess the dose-dependent effects of histamine analogs and L-histidine on mice within a social interaction apparatus to understand their impact on social behavior. Additionally, we will explore the effects of histamine antagonists on social interaction in mice. To complement these behavioral analyses, we will utilize an actophotometer to examine the movement patterns of mice and study locomotor activity. Through these approaches, we seek to gain a comprehensive understanding of how histaminergic signaling influences social behavior in mice.

#### **MATERIALS AND METHODS**

In this study, male Swiss albino mice (25-30 g) of either sex were obtained from Jawaharlal Nehru Cancer Hospital and Research Centre in Bhopal. They were housed in acrylic cages (24 x 17 x 12 cm) under controlled environmental conditions (24  $\pm$  1°C, 50  $\pm$  2% RH), maintained on a 12:12 h light/dark cycle (lights on at 07:19:00h), and provided standard diet and water ad libitum. All experimental procedures were conducted following ethical guidelines approved by the Committee for the Control and Supervision of Experimental Animals (CCSEA) and the Institutional Animal Ethics Committee of Institute SIRT Pharmacy.

#### **Drug and Solution**

L-histidine was obtained from Bioven Ingredients, while histamine receptor antagonists, cetrizine (H1 receptor antagonist), and ranitidine hydrochloride (H2 receptor antagonist) were generously provided by Bioven Ingredients. All drugs were dissolved in 0.9% saline solution. The doses used were determined based on preliminary results and literature review. For intracerebroventricular (i.c.v.) administration, aCSF solution was employed, and drugs were administered at 5 µl/mice. For intraperitoneal (i.p.) administration, a dose of 10 ml/kg was used.

# **Intracerebroventricular** (i.c.v.) **Cannulation**

Mice underwent i.c.v. cannulation under anesthesia induced by a combination of ketamine (100 mg/kg, s.c.) and xylazine (5 mg/kg, s.c.). A guide cannula (24 gauge) was stereotactically implanted according to coordinates referenced from Paxinos and Franklin. After fixation with mounting screws and dental cement, a stainless steel dummy cannula was inserted to block the guide cannula when not in use. Post-surgery, mice received cefotaxime (50 mg/kg/day, subcutaneous) for 1 week for recovery and habituation to experimental protocols. Injections were performed using a Hamilton syringe connected to an internal cannula (41 gauge), with a volume of 1.0 μL injected into the right ventricle over 2 minutes (Passani *et al.,* 2011).

#### **Sociability Test**

The sociability test was conducted in a threechambered device made of transparent plexiglass. Experimental mice were habituated for 5 minutes in a central room with closed doors to both side rooms, one containing an adult male stranger (Stranger 1) and the other an inanimate object. The doors were then opened, allowing mice to explore for 10 minutes. Time spent interacting with Stranger 1 was recorded. Subsequently, mice were subjected to a social novelty test, wherein they were acclimated for 10 minutes in the presence of two unfamiliar mice (one on each side) in a central chamber, with the second unfamiliar mouse acting as the novel stimulus (Stranger 2). Time spent interacting with Stranger 2 was recorded during a 10 minute exploration period (Yoshikawa *et al.,*  2006).



#### *Equipment and room setup*

The equipment setup for the sociability and social novelty preference tests, based on Crawley's method, consists of a threechambered rectangular box, each chamber measuring 19 x 45 cm, with clear plexiglass septa allowing free access. Two wire containers, each housing an unfamiliar mouse, are placed vertically in the side chambers. Behavioral testing is conducted between 10:00 am and 5:00 pm, with general room lighting maintained at 650 lux. Chambers are cleaned with 70% ethanol between trials for proper disinfection. Observations are made from a distance of at least 2 meters from the apparatus to minimize interference (Lee *et al.,*  2005).

#### **Habituation (Adaptation)**

To acclimate the mice to the testing environment, a plexiglass partition wall is used to separate the left and right chambers. Empty wire cages are placed in the center of the left and right chambers, with one cage for each side. The experimental mice are placed in the center of the central chamber for a 5 minute habituation period (Takahashi *et al.,*  2006).

### **Social Affiliation Aspect of the Test (Session I)**

During this session, a control mouse ("Stranger 1") is placed in a wire cup in one of the side compartments, with the placement alternating systematically between trials. The walls between the chambers are removed, allowing the experimental mice to freely explore all three chambers. The following parameters are monitored and recorded:

Number and duration of direct interactions between the experimental mice and the cup containing or without the Stranger 1 mouse, separately for each chamber. Direct contact or stretching of the experimental mice's body within a 1 cm radius around the cup is considered as active contact. Duration and frequency of other behaviors exhibited by the experimental mice in each compartment, including walking, grooming, freezing (not moving the body for more than 5 seconds), jumping, and repetitive actions. Duration and frequency of entries into each compartment, with a mouse considered inside a chamber when its head and four paws are within. Session I lasts for a duration of 10 minutes (Umathe *et al.,* 2008).

# **Social Novelty/Preference Session of the Test (Session II):**

In this session, a second control mouse ("Stranger 2") is placed in an identical wire cup in the opposite compartment, which was empty during Session I. The same parameters as described in the first session are monitored to differentiate the behavior of the experimental mice in the presence of Stranger 1 compared to Stranger 2. Session II also lasts for 10 minutes.

# **Experimental Design**

# **Effect of Histamine Precursor L-histidine on Sociability Test:**

Separate groups of mice (n=6) received intraperitoneal (i.p.) injections of either vehicle (10 ml/kg) or L-histidine (500 or 1000 mg/kg). Since the maximum effect of 500 or 1000 mg/kg of L-histidine occurred 2 hours post-treatment, mice were treated with Lhistidine 2 hours before the test. After 20 minutes, mice were subjected to the

sociability test apparatus for 10 minutes to assess sociability and social novelty-seeking behavior (Moy *et al.,* 2004).

# **Effect of H1 and H2 Receptor Antagonists on Sociability Test:**

In separate groups  $(n=6)$ , mice were intracerebroventricularly (i.c.v.) treated with either artificial cerebrospinal fluid (aCSF) (5 µl/mice), H1 receptor antagonist cetirizine (20 µg/mice), or H2 receptor antagonist ranitidine (20 µg/mice). After 20 minutes, mice underwent the sociability test for 10 minutes to evaluate sociability and social noveltyseeking behavior.

# **Effect of H1 or H2 Receptor Antagonists on L-histidine-induced Effects on Sociability Test:**

Separate groups of mice (n=6) received i.c.v. administration of aCSF (5 µl/mice), H1 receptor antagonist cetirizine (20 µg/mice), or H2 receptor antagonist ranitidine (20 µg/mice) after 1 hour and 45 minutes of i.p. injection of either vehicle (10 ml/kg) or Lhistidine (1000 mg/kg). After 20 minutes, mice underwent the sociability test for 10 minutes to assess sociability and social novelty-seeking behavior.

# **Effect of Histaminergic Analogs on Locomotor Activity:**

Different groups of mice received injections of vehicle (10 ml/kg, i.p.), L-histidine (10  $\mu$ g/mice, i.c.v or 1000 mg/kg, i.p.), H1 receptor antagonist cetirizine (20 µg/mice, i.c.v.), H2 receptor antagonist ranitidine (20 µg/mice, i.c.v.), or a combination of both antagonists with L-histidine (1000 mg/kg, i.p.). After appropriate time periods, mice were subjected to the sociability test apparatus

for 10 minutes to evaluate sociability and social novelty-seeking behavior (Crawley; 2007).

#### **Statistical Analysis**

Data are presented as mean  $\pm$  SEM. One-way analysis of variance (ANOVA) was used for most data analyses. Repeated-measures ANOVA was applied to analyze data from the sociability apparatus and locomotor activity trials. Newman-Keuls or Tukey's multiple comparison tests were used as follow-up tests. A significance level of  $P < 0.05$  was considered statistically significant. Data analysis was performed using Prism Pad Chart.

#### **RESULTS AND DISCUSSION**

### **Effect of histamine precursor L-histidine in sociability test**

In the sociability test, the effects of Lhistidine treatment at doses of 500 and 1000 mg/kg (i.p.) were evaluated using one-way ANOVA, revealing a significant impact [F(6, 35) = 39.93, \*P < 0.0001]. Newman-Keuls test indicated a significant increase in sociability with L-histidine at 1000 mg/kg (i.p.) compared to saline-treated mice ( ${}^*P$  < 0.001). However, the 500 mg/kg dose showed no significant effect ( $P < 0.05$ ). Histamine precursor L-histidine potentiates the Histamine H1 receptor subunit, influencing social behavior. Histamine synthesis can be enhanced by L-histidine administration, as seen in increased histamine levels in mice after 1000 mg/kg i.p. administration. Modulating endogenous histamine levels may offer a novel approach to controlling social behavior. Additionally, histamine H1 receptor antagonism in mice reduces activity,

potentially influencing serotonin levels in the brain.

# **Effects of H1 receptor antagonist and H2 receptor antagonist on sociability test**

The effects of H1 receptor antagonist cetirizine (20 µg/mice; i.c.v.) and H2 receptor antagonist ranitidine (20 µg/mice; i.c.v.) on the sociability test were evaluated through one-way ANOVA, revealing significant impacts  $[F(2, 15) = 61.06, *P <$ 0.0001]. Newman-Keuls test indicated significant reductions in sociability with both cetirizine and ranitidine compared to aCSF-treated mice (\*P < 0.001). Furthermore, histamine H2 receptor antagonism and H1 receptor agonism enhance serotonin release, suggesting an enhancement of social interaction through H1 and H2 receptor-mediated serotonin enhancement.

To further investigate this hypothesis, the effects of histamine H1 and H2 receptor antagonists on L-histidine-induced social ability were explored. It was found that central administration of H2 receptor antagonist ranitidine blocked the increase in social interaction induced by L-histidine, while H1 receptor antagonist showed no effect. These findings support the hypothesis that enhancing central histaminergic transmission with L-histidine leads to increased serotonin levels, further promoting histaminergic transmission.

Additionally, blocking histamine H2 receptors reduces task performance in mice, indicating a potential role in exercise-related actions. The study also detected increased social interaction after administration of H1 and H2 receptor antagonists, suggesting their potential utility in relieving negative symptoms such as social withdrawal in schizophrenia and psychotic disorders.

# **Effect of H1 and H2 receptor antagonist on L-histidine induced effect on sociability test**

The effects of H1 and H2 receptor antagonists on L-histidine-induced sociability test were examined through one-way ANOVA, revealing significant impacts  $[F(3, 20) =$ 51.16, \*P < 0.0001]. Post hoc Newman-Keuls test showed that H2 receptor antagonist ranitidine (20 µg/mice; i.c.v.) reversed the decrease in social interaction induced by Lhistidine (\*P < 0.001). However, H1 receptor antagonist cetirizine (20 µg/mice; i.c.v.) failed to affect L-histidine-induced effects on the sociability test ( ${}^*P > 0.05$ ). The study further suggests an interaction between histamine and serotonin systems, as evidenced by an increase in histamine levels upon local serotonin infusion in the hypothalamus, attenuated by a 5-HT2C receptor antagonist. Anxiety disorders, believed to stem from serotonin 5-HT2C receptor activation, are influenced by serotonin agonists like mCPP, which modulate animal behavior via the 5- HT2C receptor subtype. The reduction in social behavior after enhancing central histaminergic transmission with histamine or L-histidine may be attributed to increased serotonin levels, further augmenting histaminergic transmission via 5-HT2C/2A receptor activation. These findings suggest potential interactions between histamine and serotonin in modulating social behavior, warranting further investigation with specific serotonin analogues. However, some researchers argue that the sociability test may not effectively differentiate between anxiolytic and anticompulsive agents.

## **Effect of histaminergic analogues on locomotor activity**

Various histaminergic analogues were tested for their effect on locomotor activity at the doses as described previously using actophotometer. Table 1 illustrates the effects of various histaminergic analogues on locomotor activity in mice. Baseline locomotor activity readings were taken before treatment, and subsequent readings were recorded after treatment administration. Histamine administration via intracerebroventricular (i.c.v.) route at doses of 0.5 µg/mice and 10 µg/mice resulted in slight increases in locomotor activity compared to baseline. Conversely, L-histidine administration at 10 µg/mice via i.c.v. route led to a modest decrease in locomotor activity, while at a higher dose of 1000 mg/kg via intraperitoneal (i.p.) route, it caused a significant increase.

Treatment with the H1 receptor antagonist cetirizine (20 µg/mice; i.c.v.) slightly elevated locomotor activity, whereas treatment with the H2 receptor antagonist ranitidine (20 µg/mice; i.c.v.) led to a slight decrease. Interestingly, the combination of cetirizine or ranitidine with L-histidine (1000 mg/kg; i.p.) resulted in an increase in locomotor activity compared to baseline. The results suggest that histaminergic analogues can modulate locomotor activity in mice, with histamine and L-histidine exerting contrasting effects depending on the dose and route of administration. Moreover, the combination of histamine receptor antagonists with Lhistidine appears to influence locomotor activity differently than either compound alone.



**Figure 1: Effects of histamine, histamine precursor; L-histidine on sociability test**



**Figure 2: Effects of H1 receptor antagonist and H2 receptor antagonist on sociability test**



# **Figure 3: Effects of H1 or H2 receptor antagonist on L-histidine induced decrease in sociability test**





#### **CONCLUSION**

In conclusion, our study elucidates the significant role of histaminergic transmission in modulating social interaction and locomotor activity in mice. We found that histamine precursor L-histidine enhances sociability, while histamine receptor antagonists, particularly H1 and H2 receptor antagonists, reduce social behavior. Moreover, the combined administration of Lhistidine with histamine receptor antagonists alters sociability differently, indicating complex interactions within the histaminergic system. Additionally, histaminergic analogues exert diverse effects on locomotor activity, with implications for understanding motor behavior regulation. Overall, these findings underscore the intricate involvement of histaminergic signaling in regulating both social behavior and locomotion, providing insights into potential therapeutic strategies for conditions involving social dysfunction and motor impairments.

#### **DECLARATION OF INTEREST**

The authors declare no conflicts of interests. The authors alone are responsible for the content and writing of this article.

#### **REFERENCES**

- $\bullet$
- Akash, Y. & Dinesh, J. (2011) Formulation and evaluation of mucoadhesive microspheres of propanolol hydrochloride for sustained drug delivery. *Asian Journal of Pharmacy and Medical Sciences*, 1, 1– 8.
- Albert, A.A. & Serjeant, E.P. (1984). *Ionization Constants of Acids and Bases*. Wiley: New York, USA.
- Albrecht, K., Zirm, E.J., Palmberger, T.F., Schlocker, W. & Bernkop-Schnürch, A. (2006) Preparation of thiomer microparticles and in vitro evaluation of parameters influencing their mucoadhesive properties. *Drug Development and Industrial Pharmacy*, 32, 1149–1157.
- Amit, N.K., Ruma, M. & Biswarup, D. (2010) Gastroretentive drug delivery systems: A review. *Asian Journal of Pharmaceutical and Clinical Research*, 3, 2–10.
- Bardonnet, P.L., Faivre, V., Pugh, W.J., Piffaretti, J.C. & Falson, F. (2006) Gastroretentive dosage forms: Overview and special case of Helicobacter pylori. *Journal of Controlled Release*, 111, 1–18.
- Berthold, A., Cremer, K. & Kreuter, J. (1996) Influence of crosslinking on the acid stability and physicochemical properties of chitosan microspheres. *STP Pharma Sciences*, 6, 358–364.
- Brahmaiah (2013) B,Desu Kumar Prasanna,Nama Sreekanth, Khalilullah sd. Babu S. Satish,formulation and evaluation of extended release mucoadhesion microspheres of simvastatin. *International Journal of Pharmaceutical and Biomedical Research*, 4, 57–64.
- Brannon-Peppas, L.B. & Peppas, N.A. (1989) Solute and penetrant diffusion in swellable polymers. IX. The mechanisms of drug release from pHsensitive swelling-controlled systems. *Journal of Controlled Release*, 8, 267– 274.
- Bravo-Osuna, I., Vauthier, C., Farabollini, A., Palmieri, G.F. & Ponchel, G. (2007) Mucoadhesion mechanism of chitosan and thiolated chitosan-poly(isobutyl cyanoacrylate) core-shell nanoparticles. *Biomaterials*, 28, 2233–2243.
- Chaturvedi, G. & Saha, R. (2009). *A Review on Microsphere Technology and Its Application*. Birla Institute of Technology and Sciences, pp. 56–58.
- Chien, Y.W. (1992) Novel drug delivery systems. *Drugs and the Pharmaceutical Sciences*, 50.
- Dawood, N.M., Abdal-Hammid, S.N. & Hussien, A.A. (2018) Formulation and characterization of lafutidine nanosuspension for oral drug delivery system. *International Journal of Applied Pharmaceutics*, 10, 20–30.
- Deshmukh, R., Harwansh, R.K., Prajapati, M. & Sharma, B. (2023) Formulation and evaluation of oral mucoadhesive microspheres of ofloxacin for peptic ulcer use. *Trends in Sciences*, 20, 5751–58.
- Dewangan, H.K., Sharma, A., Mishra, A. & Singour, P. (2021) Mucoadhesive microspheres of atorvastatin calcium: Rational design, evaluation and enhancement of bioavailability. *Microscopy*, 19, 20.
- Dhanaraju, M.D., Mani Kumar, R., Nithya, P., Kishan, J.V.N. & Thirumurugan, G. (2009) Controlled delivery of antiretroviral drug loaded chitosan cross linked microspheres. *Archives of Applied Science Research*, 1, 279–286.
- Dolas, R.T., Dr Sharma, S. & Sharma, M. (2018) Formulation and evaluation of gastroretentive floating tablets of lafutidine. *Journal of Drug Delivery and Therapeutics*, 8, 393–399.
- Dorofeev, V.L., Arzamastsev, A.P. & Veselova, O.M. (2004) Melting point determination for the analysis of drugs of the fluoroquinolone group. *Pharmaceutical Chemistry Journal*, 38, 333–335.
- *European Pharmacopoeia* (2004), Vol. 1. Directorate for the Quality of Medicines of the Council of Europe (EDQM), p. 628.
- Fefelova, N., Nurkeeva, Z., Mun, G. & Khutoryanskiy, V. (2007) Mucoadhesive interactions of amphiphilic cationic copolymers based on  $[2]$ (methacryloyloxy)ethyl]trimethylamm onium chloride. *International Journal of Pharmacy*, 33, 25–32.
- Ghumman, S.A., Noreen, S. & Tul Muntaha, S. (2020) Linum usitatissimum seed mucilage-alginate mucoadhesive microspheres of metformin HCl: Fabrication, characterization and evaluation. *International Journal of Biological Macromolecules*, 155, 358–368.
- Gulia, Y. & Choudhary, M.  $(2011)$ Peptic ulcer disease: A review. *Pharmacologyonline*, 3, 48–70.
- Haas, J. & Lehr, C.M. (2002) Developments in the area of bioadhesive drug delivery systems. *Expert Opinion on Biological Therapy*, 2, 287–298.
- Higuchi, T. (1963) Mechanism of sustained-action medication: Theoretical analysis of rate of release of solid drugs dispersed in solid matrices. *Journal of Pharmaceutical Sciences*, 52, 1145–1149.
- Huang, J.Q., Sridhar, S. & Hunt, R.H. (2002) Role of Helicobacter pylori infection and non-steroidal antiinflammatory drugs in peptic-ulcer disease: A meta-analysis. *Lancet*, 359, 14–22.
- *Indian Pharmacopeia* (2007), Vol. 2.
- Kaur, A., Singh, R., Sharma, R., Kumar, S. & Ulcer, P. (2012) A review on Ethology and pathogenesis. *International Research Journal of Pharmacy*, 8407, 2230.
- Kavita, D. & Piyush, T.  $(2013)$ Formulation and evaluation of mucoadhesive microspheres of rantidine hydrochloride using chitosan and sodium carboxymethyl cellulose as polymers. *International Journal of Pharmaceutical and Biomedical Research*, 4, 140–144.
- Khutoryanskiy, V.V. (2007) Hydrogen-bonded interpolymer complexes as materials for pharmaceutical applications. *International Journal of Pharmaceutics*, 334, 15–26.
- Klausner, E.A., Eyal, S., Lavy, E., Friedman, M. & Hoffman, A. (2003) Novel levodopa gasrtroretentive dosage form: In vivo evaluation in dogs. *Journal of Controlled Release*, 88, 117–126.
- Korsmeyer, R.W., Gurny, R., Doelker, E., Buri, P. & Peppas, N.A. (1983)

Mechanisms of solute release from porous hydrophilic polymers. *International Journal of Pharmaceutics*, 15, 25–35.

- Kumar, S., Tiwari, A. & Goyal, N. (2022) Floating Microspheres of lafutidine: Formulation, Optimization, Characterization, in-vitro and in-vivo Floatability Studies Using Eudragit Grades. *Indian Journal of Pharmaceutical Education and Research*, 56, 681–688.
- Ludwig, A. (2005) The use of mucoadhesive polymers in ocular drug delivery. *Advanced Drug Delivery Reviews*, 57, 1595–1639.
- Nagahara, N., Akiyama, Y., Nakao, M., Tada, M., Kitano, M. & Ogawa, Y. (1998) Mucoadhesive microspheres containing amoxicillin for clearance of helicobactor pylori. *Antimicrobial Agents and Chemotherapy*, 42, 2492– 2494.
- Neha, B. & Katla, D.V. (2023) Design and characterization of Methotrextate mucoadhesive Microspheres. *International Journal of Pharmacy Research & Technology (IJPRT)*, 13, 35–45.
- Nicholls, T.J., Green, K.L., Rogers, D.J., Cook, J.D., Wolowacz, S. & Smart, J.D. (1996) Lectins in ocular drug delivery, An investigation of lectin binding sites on the corneal and conjunctival surfaces. *International Journal of Pharmaceutics*, 138, 175– 183.
- Nighute, A.B.  $&B$  Bhise, S.B. (2009) Preparation and evaluation of rifabutin loaded polymeric microspheres.

*Research Journal of Pharmacy and Technology*, 2, 371–374.

- Pantwalawalkar, J. & Nangare, S. (2022) Formulation of silk fibroinbased single polymeric floating microspheres for sustained release of lafutidine. *Indian Journal of Pharmaceutical Education and Research*, 56, 396–404.
- Patel,  $V.$  (2013) Design, development,evaluation and optimization of antiulcer delayed release tablets. *Research Journal of Pharmacy and Technology*, 6, 669– 684.
- Pattanayak, S., Kanthal, L.K., Afnan, T., Shukla, S. & Panda, P.K. (2024) Preparation and evaluation of ketoprofen loaded mucoadhesive microspheres. *Journal of Drug Delivery and Therapeutics*, 14, 4–8.
- Pawar, V.K., Kansal, S., Garg, G., Awasthi, R., Singodia, D. & Kulkarni, G.T. (2011) Gastroretentive dosage forms: A review with special emphasis on floating drug delivery systems. *Drug Delivery*, 18, 97–110.
- Pendyala, Y. & Talasila, S. (2012) Formulation and evaluation of chitosan loaded mucoadhesive microsphere of ramipril. *International Journal of Pharmaceutical and Chemical Sciences*, 1, 904–911.
- Priyadarshini, M.K., Parthiban, S., Senthil Kumar, G.P. & Tamizh Mani, T. (2014) Preparation and evaluation of microspheres encapsulating zidovudine. *Int. J. Res Pharma and Nano Sci.*, 3, 461–468.
- Radha.G.V,Sraranthi.N.Lakshmi,Swet ha.P,K.Sravani,Kumar.K.Parveen,For mulation and evaluation of mucoadhesive microspheres of nifedipine,Journal of Pharmaceutical and Scientific Innovation (2012), 1, 39–43.
- Raghavendra Rao, N.G.R. & Bisht, A. (2021) Formulation and evaluation of lafutidine gas powered system for controlled release. *International Journal of Pharmaceutical Sciences Review and Research*, 66, 119–125, Article no. 19.
- Rao, V.B., Schaitanya, K. & Kumar, K.A. (2012) Allamneni Yaswanth Reddy B.V.V.V.K, chary. P. Dayananda, N performance evaluation of mucoadhesive microspheres containing an antidiabetic drug: Glipizide. *International Journal of Pharmaceutical Sciences and Drug Research*, 4, 115–122.
- Sagar, S. & Pramodini, G.N. (2023) formulation development and characterization of lafutidine raft system. *International Journal of Pharmacy and Pharmaceutical Sciences*, 15, 8–15.
- Salman, A., Tsror, L., Pomerantz, A., Moreh, R., Mordechai, S. & Huleihel, M. (2010) FTIR spectroscopy for detection and identification of fungal phytopathogenes. *Spectroscopy*, 24, 261–267.
- Shailaja, P., Ashutosh, B. & Kothiyal, P. (2016) A review on gastroretentive drug delivery system. *Int. J. Res. Dev. Pharm. L. Sci.*, 5, 2178–2187.
- Sharma, M., Pk, C. & Dev, S.K. (2017) FORMULATION AND IN-VITRO-IN-VIVO EVALUATION OF ALGINATE-CHITOSAN MICROSPHERES OF GLIPIZIDE BY IONIC GELATION METHOD. *Asian Journal of Pharmaceutical and Clinical Research*, 10, 385–390.
- Shivhare, U.D. & Tijare, P.M.  $(2013)$ Formulation and characterization of microspheres of selected anti-infective agent for urinary tract infection. *Journal of Drug Delivery Research*, 2, 16–26.
- Streubel, A., Siepmann, J. & Bodmeier, R. (2006) Gastro retentive drug delivery system. *Expert Opinion on Drug Delivery*, 3, 217–233.
- Takenaka, R., Okada, H., Kawano, S., Komazawa, Y., Yoshinaga, F., Nagata, S., Inoue, M., Komatsu, H., Onogawa, S., Kushiyama, Y., Mukai, S., Todo, H., Okanobu, H., Manabe, N., Tanaka, S., Haruma, K. & Kinoshita, Y. (2016) Randomized study of lafutidine vs lansoprazole in patients with mild gastroesophageal reflux disease. *World Journal of Gastroenterology*, 22, 5430–5435.
- Teli, L.K.S., Tyagi, Y. & Raghavendra Rao, N.G. (2020) Formulation and evaluation of lafutidine loaded fast dissolving tablet using Trigonella foenum. *Indian Journal of Research in Pharmacy and Biotechnology (IJRPB)*, 8, 1–9.
- Thejeswini, K., Sowmya, C., Sunitha, J. & Surekha, R. (2014) Formulation development and evaluation of microspheres containing lopinavir. *Int.*

*J. Innovative PharmSci Res*, 2, 1638– 1648.

- Truter, I. (2009) Evidence-based pharmacy practice (EBPP): Peptic ulcer disease. *South African Pharmaceutical Journal*, 76, 10–20.
- Umbare, R.P., Mate, G.S. & Dongare, S.S. (2011) Antiulcer activity of crude alcoholic extract of rhizomes of Cissus repens Lan. *Research Journal of Pharmacy and Technology*, 4, 60–62.
- Vimal, Y.  $(2011)$  K, Kumar Brajesh,Prajapati.S.K,Shafaat Kausar,Design and evaluation of mucoadhesive microspheres of repaglinide for oral controlled releas. *International Journal of Drug Delivery*, 3, 357–370.
- Yalkowiski, S.H. & Roseman, T.J. (1981), Chapter 3. *Techniques of Solubilisation of Drugs* (edited by S. H. Yalkowski, M. Dekker & Newy).
- Yie, W.C. (1992). *Concepts and System Design for Rate Controlled Drug Delivery in Novel Drug Delivery System*,  $2^{nd}$  edn. Marcel Dekker, Inc.: New York, USA.